STUDY PROTOCOL

Modifying Intestinal Integrity and MicroBiome in Severe Malnutrition with Legume-Based Feeds (MIMBLE 2.0): protocol for a phase II refined feed and intervention trial [version 1; peer review: 2 approved]

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Abstract

Background: Changes in intestinal mucosal integrity and gut microbial balance occur in severe acute malnutrition (SAM), resulting in treatment failure and adverse clinical outcomes (gram-negative sepsis, diarrhoea and high case-fatality). Transient lactose intolerance, due to loss of intestinal brush border lactase, also complicates SAM, thus milk based feeds may not be optimal for nutritional rehabilitation. Since the gut epithelial barrier can be supported by short chain fatty acids, derived from microbiota fermentation by particular fermentable carbohydrates, we postulated that an energy-dense nutritional feed comprising of legume-based fermentable carbohydrates, incorporated with lactose-free versions of standard World Health Organization (WHO) F75/F100 nutritional feeds will enhance epithelial barrier function in malnourished children, reduce and promote resolution of diarrhoea and improve overall outcome.

Methods: We will investigate in an open-label trial in 160 Ugandan children with SAM, defined by mid-upper arm circumference <11.5cm and/or presence of kwashiorkor. Children will be randomised to a
lactose-free, chickpea-enriched feed containing 2 kcal/ml, provided in quantities to match usual energy provision (experimental) or WHO standard treatment F75 (0.75 kcal/ml) and F100 (1 kcal/ml) feeds on a 1:1 basis, conducted at Mbale Regional Referral Hospital nutritional rehabilitation unit. The primary outcomes are change in MUAC at day 90 and survival to day 90. Secondary outcomes include: i) moderate to good weight gain (>5 g/kg/day), ii) de novo development of diarrhoea (>3 loose stools/day), iii) time to diarrhoea resolution (if >3 loose stools/day), and iv) time to oedema resolution (if kwashiorkor) and change in intestinal biomarkers (faecal calprotectin).

**Discussion:** We hypothesize that, if introduced early in the management of malnutrition, such lactose-free, fermentable carbohydrate-based feeds, could safely and cheaply improve global outcome by reducing lactose intolerance-related diarrhoea, improving mucosal integrity and enhancing immunity, and limiting the risk of systemic infection and associated broad-spectrum antibiotic resistance.

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**Keywords**
severe malnutrition, nutritional feeds, African children, gut-barrier dysfunction, legume-based fermentable carbohydrates F75 F100 clinical trial, Uganda

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Introduction
Severe acute malnutrition (SAM) accounts for 1.5 million deaths worldwide annually\(^1\). Undernutrition, highlighted in a Lancet series\(^2\), is a greatly neglected area of research despite contributing to around 60% of childhood deaths. Community-based care with ready-to-use therapeutic food for children with uncomplicated malnutrition has markedly improved recovery rates; but little progress has been made for children hospitalised with SAM in whom mortality remains high\(^1\). Recommended nutritional support has shown improvements in growth but these often do not translate to improving mortality and relapse rates, indicating that current strategies are not working in practice. In order to understand why current nutritional feeds have limited impact on longer-term mortality rates despite nutritional recovery, we need a new approach to understanding the complex interaction between nutrition, the gut and its associated microbiome. For example, in African children diarrhoea complicates 65% of these children and is associated with high morbidity and mortality rates (20%)\(^3-5\). SAM causes disruption of normal intestinal flora\(^6\), increased turnover of vital nutrients, disruption of gut barrier function, impaired mucosal immunity and increased risk of gram negative bacteraemia\(^7\), which together form a vicious cycle\(^8\).

Lactose intolerance in SAM
The current recommended formulae by WHO, initially F75 followed by F100, are milk-based feeds and their major carbohydrate source are disaccharides (a mixture of maltodextrin, sucrose and lactose), which can cause osmotic diarrhoea since they normally are hydrolysed by disaccharidases localized at the tip of small intestinal villi. Both lactose intolerance\(^9\) and intestinal atrophy have been demonstrated in children with SAM\(^10-12\). Removing lactose and replacing it with better-tolerated simple carbohydrates that do not rely on brush border enzymatic activity for digestion and absorption may be highly beneficial in preventing and ameliorating diarrhoea in SAM.

Benefits of non-digestible fermentable carbohydrates
Numerous studies have shown that intestinal mucosal integrity and gut microbial balance can be restored by inducing fermentation in the gastrointestinal tract\(^13\). Fermentable carbohydrates are increasingly being investigated as potential adjuncts to improve the balance of normal gut flora and positively influence the immunological and metabolic function of the gut\(^14\). Non-digestible fermentable oligosaccharides induce favourable colonic microbiota fermentation\(^15\), leading to the generation of short-chain fatty acids (SCFA), which have a positive influence on gut integrity and nutritional health by improving energy yield, production of vitamins and the stimulation of gut homeostasis, including anti-pathogen activities\(^16\)\(^17\). Preliminary data collected from children with SAM by a multi-disciplinary team at Imperial College (SMIP study) involving specialists in international child health (K.M.), nutrition (G.F.), metabonomics (E.H.) and colonic bacterial metataxonomics (J.M.) suggest that there are changes in the inter-relationships of normal gut microbes, intestinal permeability, and markers of gut barrier dysfunction and immune dysregulation in SAM which effect clinical outcome (manuscript in preparation) and are therefore potential targets for intervention.

In the MIMBLe (Modifying Intestinal Integrity and MicroBiome in Severe Malnutrition with Legume-Based Feeds) pilot study we tested a locally made cowpea flour using a “Posho” mill, which was added to standard nutritional feeds (as in Figure 1) (Pan African Clinical Trial Registry: PACTR201805003381361). This small study demonstrated that the feed is well tolerated when added to the nutritional recovery programme, and increased bacterial diversity compared to standard feeds. We also demonstrated decreases in anorectic gut hormones (manuscript in preparation).

Sources of non-digestible fermentable carbohydrate
Legumes, which contain a mixture of different types of non-digestible fermentable carbohydrates, have been demonstrated to augment microbial communities that enhance SCFA production\(^18\). Furthermore, these legumes form part of the traditional East African diet consumed by children\(^9\) and a number of current research programmes in East Africa are expanding legume growth to improve the environmental impact of agriculture (nitrogen fixing)\(^19\), meaning that this treatment will be both acceptable and readily available to local communities. One potential commonly consumed food with prebiotic effect is chickpea. Chickpea-based follow-on formulae have been explored as a potential prevention for undernutrition\(^20\).

Summary and importance
There are multiple reasons why nutritional support, at an early stage in the management of SAM, tailored towards prevention of diarrhoea and restoring gut mucosal integrity and manipulation of the gut microbiome could enhance resilience to secondary infection and morbidity during re-feeding. Legumes, through the support of microbiota networks to produce short chain fatty acids, offer a safe therapeutic adjuvant that is easily and cheaply sourced in East Africa, and other regions with a high burden of severe malnutrition. Additional removal of lactose from therapeutic
feeds should reduce incidence of osmotic diarrhoea, and related morbidity.

The planned research will bring new knowledge to the field of severe malnutrition, thereby addressing a major treatment gap, which is a high priority research issue and a significant barrier to progress in the reduction of mortality. The study has the potential to suggest new therapies for enhancing recovery of gut mucosal function and resilience to infection through the modulation of the gut microbiome, thereby reducing mortality and morbidity in SAM. As the interventional feed from the original MIMBLE pilot is being refined in conjunction with a specialist food company (Campden BRI), this offers an opportunity to improve the food safety profile, product utility, and further optimise the feed by removing lactose.

Study protocol
Justification for the study
This phase II study will be the first step in evaluating whether giving children with severe acute malnutrition an altered nutritional feed helps repair the gut and encourage the healthy microbiome, which may reduce illness or improve recovery. In the experimental arm (legume-based feed) the standard nutritional feed that all children with severe malnutrition receive routinely will be altered by removal of lactose and adding a chickpea flour. Chickpea flour contains fibre/roughage that can help encourage SCFA production leading to modulation of the gut microbiome, thereby reducing mortality and morbidity in SAM.

Hypothesis
We hypothesize that legume-enriched, lactose-free nutrition support will improve outcomes in children with severe acute malnutrition, compared with standard nutritional rehabilitation milks as recommended by the WHO.

Objectives
To generate additional proof supporting the hypothesis that non-digestible fermentable carbohydrate in the form of chickpea flour will have positive effects on intestinal mucosal integrity and permeability, gut inflammation, gut microbiome and clinical outcomes in children with severe acute malnutrition compared to children receiving standard nutritional feeds. Also to provide evidence that a lactose-free feed will reduce the incidence and encourage the resolution of diarrhoea, which is a major complicating factor in SAM.

Design and methodology
The study will be conducted in Mbale Regional Referral Hospital (MRRH), Uganda. MIMBLE 2 progress at the time of writing is detailed in the ‘Study progress’ section below.

Study design
A single-centre, open-label pilot phase II trial comparing a lactose-free, chickpea enriched feed containing 2 kcal/ml, provided in quantities to match usual energy provision (experimental) to WHO standard treatment F75 (0.75kcal/ml) and F100 (1kcal/ml) feeds randomised on a 1:1 basis.

Study location
The study will be conducted at the Paediatric Nutrition Unit at Mbale Regional Referral Hospital, Eastern Uganda. The 17-bedded nutrition unit in Mbale admits 300 children each year with severe acute malnutrition. Most of the clinical services provided by the paediatric department are free of charge to the patients in accordance with the government policy of no cost sharing in public hospitals.

Study population
160 children aged > 6 months to 5 years. Inclusion criteria: signs of severe malnutrition one of (i) Marasmus (defined by mid-upper arm circumference (MUAC) <11.5cm) and/or Kwashiorkor (defined as symmetrical pitting oedema involving at least the feet irrespective of WHZ score or MUAC) and a guardian or parent willing/able to provide consent. Exclusion criteria: children with severe acute malnutrition with a very high risk of death due to a comorbidity e.g. malignant disease or terminal illness will be excluded.

Sample size
Mid-upper arm circumference (MUAC) has been selected as the primary criterion for nutritional recovery because it predicts mortality better and is less affected by oedema than other anthropometric measures, and is also a good index of muscle mass. A recent trial of antimicrobial prophylaxis in Kenyan children admitted with severe malnutrition baseline mean MUAC was 10.6 cm (SD 1.06) and at 90 days, 12.2 cm (SD 1.35), a mean change of 1.6 cm (SD 1.1) nutritional recovery at 90 days. The overall sample size will be 160 children which will include 80 in each study arm. From the original MIMBLE trial, safety in the study arms were comparable to the standard intervention (manuscript in preparation). As a Phase II study, this study is designed to provide adequate proof of principal that the modified nutritional feed provides clinical, physiological and biological evidence of benefit to the child in terms of nutritional rehabilitation and the predicted effect on biological markers of gut-barrier dysfunction and on the intestinal microbiota. Additionally, it will compare the safety and tolerability of the modified nutritional feed against the current standard of care. The data will thus inform the design and effect size for future pragmatic trials. Formal sample sizes were not calculated; since the study was designed to provide safety data and some indication of the likely efficacy of the modified feed compared to standard of care. We thus aimed to study 160 children in total, which is realistic given the timeframe and funding available for the study.

Study methods and procedures
Screening procedure. Dedicated trial research nurses will be employed to conduct the study. Eligible children will be identified by the nurse and clinician on duty and registered in the eligibility screening log. A member of the trial team will then perform a structured clinical assessment of WHO anthropometric and clinical criteria to confirm severe acute malnutrition and any exclusion criteria.
Details of those fulfilling the entry criteria will be entered onto a screening form, while reasons for non-eligibility will be added to the eligibility screening log. It is anticipated that this process will take approximately 10 minutes.

Consent. Prospective written, informed consent will be sought from parents or guardians of children. Parents or guardians will be given an information sheet in their usual language containing details of this pilot study. The sheet will be read aloud to those who are unable to read. Parents and guardians will be encouraged to ask questions about the trial prior to signing the consent form (Supplementary File 1). For parents who are unable to sign the consent form, a thumbprint will be required and a witness will be required to sign the consent form. Supplementary File 2 contains the information given to the parents/guardians of each child in the study.

Randomisation. Randomisation is in permuted blocks. The sequence was generated by a data manager at KEMRI Wellcome Trust Research Programme (KWTRP), Kilifi, Kenya. This sequence was used, by a study administrator, to prepare cards with treatment allocation (standard or altered standard with chickpea flour) at the Clinical Trial Facility, KWTRP, Kilifi, Kenya. The randomisation cards were sent to MRCI, Mbale. The cards will be kept in numbered, sealed opaque envelopes, each signed across the seal. The randomisation cards are numbered consecutively and the dedicated MIMBLE clinical trial team will select the next card in the sequence ie cards will be opened in numerical order when enrolling a patient in the clinical trial.

Study interventions. Children will be randomly allocated on a 1:1 basis to:

- Control arm: World Health Organization recommended F75 (0.75 kcal/ml) and F100 (1 kcal/ml) feeds
- Experimental arm: Lactose-free, chickpea enriched feed containing 2 kcal/mL, provided in quantities to match energy provision in standard treatment.

For the control arm, children will receive the standard inpatient hospital management as per national guidelines. Briefly, children will initially receive F75 therapeutic milk at a rate of 130 ml/kg/day, provided at 4-hour intervals. When the child has stabilized (resolved hypoglycaemia, resolved hypothermia and demonstrating appetite) they will be transitioned onto F100 therapeutic milk at the same rate. The rate may be increased by 10 ml per feed, until a maximum rate of 200 ml/kg/day has been achieved. If the child is oedematous, F75 will be commenced at 130 ml/kg/day with 4-hourly feeds until stabilized (the above criteria and the resolution of oedema). When stable the child will transition to F100 formula at the same rate, and if tolerated increase to 130 ml/kg/day. If there is no indication of fluid overload, the feed may be increased by 10 ml per feed to a maximum of 200 ml/kg/day.

For the experimental arm, children will receive pre-packaged modified feeds which are lactose-free and include chickpea (gram) flour as a source of fermentable carbohydrate. The feeds contain: lactose-free skimmed milk powder (7.25%), rapeseed oil (11.5%), gram flour (10%), sugar (9%) and water. All ingredients were sourced from established EU suppliers and passed all required safety tests for human consumption as appropriate for each ingredient (contaminants, pesticides, toxins, bacterial contamination etc.). The final feed contains (per 100 ml): 200 kcal, 18 g total carbohydrate, 5.6 g protein, 12g fat, and 0.4 g resistant starch. The quantity of feed provided will match the total amounts of energy and protein that would be received in the control arm, and additional water provided to match the fluid received.

Treatment with control/experimental treatment will be for 14 days duration, followed by standard treatment outpatient as required. This includes provision of ready to use therapeutic feeds until child has recovered, defined by a weight-for-height z-score ≥2. The duration of this is variable.

Clinical management and monitoring

Clinical monitoring. Clinical monitoring will be in line with standard care at a minimum of twice daily temperature and pulse oximetry.

Anthropometric indices. Children will have admission their height or length measured (using a standard height meter or a dedicated length board) and daily weight and MUAC measurements until discharge performed at the same time each day. These measurements will be repeated on day 28 and day 90 (Table 1).

Stool assessment. Frequency and consistency of stool will be assessed daily until discharge and at day 28 and day 90.

Dietary recall. Children and/or parents/guardians will be asked to complete a 24-hour multi-pass dietary recall via a structured interview to assess the nutritional intake of children on admission, day 28 and day 90 as most children will have progressed onto normal diet with additional RUTF. Intake of standard and modified feeds will be recorded daily during the inpatient episode.

Day 28 and day 90 follow-up. Children will be invited to return to the hospital for a follow-up visit on day 28 and day 90 as a day case which will last approximately 2 hours. This will include a clinical review, anthropometric measurements, and the study investigations outlined above.

Laboratory investigations. Following consent and randomisation, routine admission blood samples will be taken, in accordance to routine clinical practice, for the following investigations: full blood count, clinical chemistry, lactate, glucose, blood culture, malaria status and HIV status. As per WHO guidance all parents will be approached to HIV testing. HIV testing will be conducted with provision of verbal parental or guardian consent. Pre- and post-test counselling will be done in accordance with routine practice. The total admission blood sampling volume equates to 10 ml, the majority of which is for clinical purposes and
Table 1. Assessment & investigation flow chart.

<table>
<thead>
<tr>
<th>Time</th>
<th>Adm</th>
<th>Day 1</th>
<th>Day 7</th>
<th>Day 28</th>
<th>Day 90</th>
<th>Discharge</th>
</tr>
</thead>
<tbody>
<tr>
<td>Consent + patient information leaflet</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Anthropometry (weight, MUAC)</td>
<td>Daily as inpatient</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Height</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Clinical observations</td>
<td>Daily as inpatient</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fluid balance</td>
<td>Daily as inpatient</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td>X</td>
</tr>
<tr>
<td>Stool Assessment</td>
<td>Daily as inpatient</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td></td>
<td></td>
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<tr>
<td>24-Dietary recall</td>
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</tr>
<tr>
<td>Length of stay</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Laboratory investigations</td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Haematology</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Biochemistry</td>
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<td>X</td>
<td>X</td>
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</tr>
<tr>
<td>Glucose</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>X</td>
</tr>
<tr>
<td>Lactate</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>X</td>
</tr>
<tr>
<td>Malaria test</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HIV testing</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Urine dipstick (Multi-stick)</td>
<td>X</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stool microscopy</td>
<td>X</td>
<td></td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

| Stored samples               |     |       |       |        |        |           |
| For Metabolomics and SCFA (NMR, GC-MS) |     |       |       |        |        |           |
| Plasma                      | X   | X     | X     | X      |        |           |
| Urine                       | X   | X     | X     | X      |        |           |
| Stool                       | X   | X     | X     | X      |        |           |
| For Metataxonomics (16s rRNA) |     |       |       |        |        |           |
| Stool                       | X   | X     | X     | X      |        |           |

| For Intestinal Cell Injury (Faecal Calprotectin) |     |       |       |        |        |           |
| Stool                                               | X   | X     | X     | X      |        |           |

only includes an extra 3 ml for plasma storage. Routine bloods will also be taken on day 7 and day 28 and day 90 which again will include an extra 3 ml for plasma storage. Plasma will be used for short chain fatty acid and metabolite analysis (Table 1).

At admission, day 7, day 28 and day 90 stool and urine samples will be taken and be stored. The total volume of urine and stool collected across the study will be 30 ml and 30 g, respectively. Stored samples will be used for metabolite analysis (urine and stool), faecal calprotectin (stool), reducing substances (stool) and DNA extraction for 16S rRNA metataxonomics (stool).

All stored samples will be sent to Imperial College, London to assess the following:

- Intestinal cell injury, by faecal calprotectin assays.
- Changes in gut microbiota, by 16S rRNA analysis for species related to gut barrier function e.g. *Bifidobacterium* and *Lactobacillus*.
- Examination of changes in host and bacterial metabolic products, by nuclear magnetic resonance spectroscopy and gas chromatography–mass spectrometry.

Standard case management: in hospital

All study patients will be otherwise managed and clinically monitored as per national guidelines; World Health Organization Guideline- Updates on the management of severe acute malnutrition in infants and children (2013). This includes management of dehydration, hypothermia, hypoglycaemia, dermatosis, treatment of infection, eye care, anaemia and vitamin and mineral supplementation. The criteria to transition from F75 to F100 will be as per national guidelines. The discharge criteria from hospital will also be as per national guidelines.

Co-primary outcome measures

1. Change in MUAC at Day 90
2. Survival to Day 90
Secondary outcomes

1. Clinical:
   a. Weight gain (moderate to good: >5 g/kg/day)
   b. De novo development of diarrhoea (>3 loose stools/day)
   c. Time to diarrhoea resolution (if >3 loose stools/day)
   d. Time to bi-pedal oedema resolution (for those with Kwashiorkor)

2. Changes in intestinal biomarkers:
   a. Intestinal cell injury (faecal calprotectin)

Supportive biological and physiological data to support end points will include:

   A. Anthropometric:
      a. Weight/height/BMI
   B. Microbiota:
      a. Percentage change in relative populations of gut microbiota
   C. Metabolomics:
      a. Changes in generation of short chain fatty acids
      b. Changes in host and microbiota metabolic products
   D. Lactose intolerance:
      a. Evidence of stool reducing substances at baseline

Safety reporting

All relevant adverse events will be reported in the case report form (CRF) and serious adverse event (SAE) forms (Supplementary File 3). The reporting procedure will be captured within a dedicated safety reporting SOP. At each clinical review the child will be assessed for adverse events, both expected and unexpected. Non-serious and expected events will be routinely captured in the CRF.

SAEs

SAEs will be reported immediately to the on-site principal investigator (PI) so that co-PIs may be notified of the event. Serious adverse events include:

1. Any untoward medical occurrence or effect that is:
   (i) fatal, (ii) life threatening, (iii) permanently or temporarily disabling or incapacitating, (iv) causes prolongation of hospital stay (if in the clinician’s judgment the adverse event causes the child to stay in hospital longer than would have been necessary if the adverse event had not occurred) or (v) any other event that investigator considers serious, having a real, not hypothetical, risk of one of the previous outcomes.

2. Allergic reactions, which will automatically be considered as SAEs in this pilot study.

A dedicated SAE form will be completed including details of the nature of event, date of onset, severity, corrective therapies given, outcome and causality (i.e. unrelated, unlikely, possible, probably, definitely) (see Table 2). This data will be reviewed as part of the study monitoring process.

Trial monitoring

This Phase II will be monitored locally, by designated GCP trained and experienced monitors, through assessment of any SAE forms and accumulation of data collected. There is no Data and Safety Monitoring Committee and no planned interim analyses.

Data management and statistical analysis

Information will be stored securely and will only be made available to those caring for the child and those directly involved in the study. Any reports will not mention the names of the child or parent/guardian. Patient admission data collection forms and observation charts will be kept in a locked cupboard. Designated members of the research team will transcribe relevant data from the source documents to the CRF. Data entry will be done through OpenClinica, an FDA-approved, web-based application, to a relational PostgreSQL database. Security is enforced

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Grade 1 mild</th>
<th>Grade 2 moderate</th>
<th>Grade 3 severe</th>
<th>Grade 4 life-threatening/fatal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clinical adverse event NOT identified elsewhere in the grading table</td>
<td>Symptoms causing no or minimal disturbance with current illness</td>
<td>Symptoms causing greater than minimal disturbance with current illness</td>
<td>Symptoms requiring corrective medical treatment</td>
<td>Symptoms requiring medical treatment to prevent permanent impairment, persistent disability, or death</td>
</tr>
<tr>
<td>Allergic reaction</td>
<td>Local urticaria or flushing with no medical intervention indicated</td>
<td>Allergic reaction: Fever, chills, flushing, limited pruritic rash, nausea and vomiting without generalised angioedema or bronchospasm</td>
<td>Grade 2 reaction PLUS Generalized urticaria OR angioedema with medical intervention indicated OR Symptomatic mild bronchospasm</td>
<td>Grade 3 reaction PLUS Acute anaphylaxis OR Life-threatening bronchospasm OR laryngeal edema</td>
</tr>
</tbody>
</table>

Table 2. Assessment of adverse event.
through authentication of users by use of encrypted passwords. Different access level accounts authorize users on actions they may perform on the database.

Data will be analysed on an intention-to-treat basis. Differences in outcome measures between the three treatment groups will be compared with ANOVA or linear regression as appropriate. Where variables are not normally distributed, they will be transformed by taking logarithms. If this is insufficient, then equivalent non-parametric tests will be used. Metabolomic data will be analysed using multivariate statistical techniques including principal components analysis, projection to latent structures (PLS) and orthogonal PLS.

Ethical statement
Ethical and regulatory approval has been granted by Imperial College Research Ethics Committee (17IC4146), the sponsor of the study and from Mbale Regional Hospital Institutional Review Committee (019/2018) and UNCST (HS2391). The MIMBLE2 study has been registered on ISRCTN (registry identifier 10309022).

Safety
There are very few risks attached to this study. Intolerance or allergy to chickpeas are rare. Details of the processing and packaging will be addressed by due diligence of the experienced collaborator (Campden BRI).

Blood samples are required as part of this study; this includes routine blood sampling according national guidelines and an additional 12 ml over the whole duration of the study. Required volumes of blood will be minimized wherever possible and be within the locally agreed maximum. Urine and stool will be collected in a non-invasive manner thus should not cause any distress to the child or their family.

Potential benefit to the child and/or family include:
• Closer observation during admission, which consequently may allow the clinical team to make-important changes to the child’s treatment during in hospital stay.
• All routine non-study medications required by the hospital to treat the child will be made available. Any additional procedure required due to a complication or a potential complication will be covered by the study.
• The parents or guardians for the children will be asked to return for follow up at 28 days after admission. This additional visit will include a clinical review thus an opportunity for medical treatment if required. Education regarding nutrition will also be readdressed at this time. Reimbursement for transport cost for this follow-up visit plus any treatment costs required during the visits will be made.

Protocol version changes
Version 1. Previous versions approved the use of locally sourced legumes which could not be certified to the same standards as those in the UK.

Version 2. First version included another recruitment site, which became unfeasible.

We do not anticipate any changes to the design of the trial; however, if there were need to alter the trial (e.g., a request for an extension in enrolment, changes or addition of a trial centre and/or investigators), this would be communicated to the ICREC, MRRHREC and UNCST via a formal request for a protocol amendment, as would any other changes in the trial protocol.

Discussion
Prior to this study we conducted a proof-of-concept study in East Africa in undernourished children MIMbLe Study (Pan African Clinical Trial Registry: PACTR201805003381361) using locally developed cowpea flour added to standard nutritional feeds. We were reassured by the safety and tolerability of this feed. In addition, cowpea-enhanced feeds resulted in a positive effect on increasing bacterial diversity and a favourable decrease in anorectic gut hormones. This led to further funding to enable product development and testing in a randomised control trial. Initially cowpea was targeted as a locally available legume that contains adequate amounts of the resistant starch of interest to promote gut microbial health. The legume-based interventions we propose are used within the local diets but have been milled/processed in such a way to maximise their ability to ferment within the bowel to create a favourable microbiome. The details of the manufacturing process and details of the final product will be available on request (manuscript in preparation).

During the 6-month period of product development we encountered two significant barriers to utilising cowpeas in the current study. First, on testing, a final cowpea-enriched feed product was found to have significantly less resistant starch than anticipated, and second we were not able to identify a commercial supplier whose products met the strict food safety standards required with respect to certification for contaminants including pesticides, toxins, microbes and metals. We therefore opted to use another locally available legume alternative, the chickpea, although this is eaten less frequently in Uganda. On testing we found that the final nutritional paste retained its resistant starch content throughout processing, moreover chickpea (gram) flour was readily available from reputable suppliers. Overall, chickpea production in Uganda in 2016 was estimated to be 5085 tonnes in 2016 by the Food and Agricultural Organization of the United Nations, increasing by approximately 100 tonnes (2.2%) each year since 2010. The safety is assured as intolerance or allergy to chick peas are rare. The G6PD variant in these populations retains >12% of its activity, thus not rendering G6PD-deficient patients to susceptibility to oxidant stress (as seen in Mediterranean variants rendering them susceptible to ‘Favism’).

Potential future impact
An increasing awareness that malnourished populations have higher levels of gut dysfunction with frequent episodes of infection, and a focus on both carbohydrate and protein requirement is inescapable. Enhancing carbohydrate delivery to the colon offers a solution to supporting a mature and diverse microbiome whilst simultaneously promoting gut integrity through SCFA
production, together with improving plant-based protein digestibility and bioavailability are both relevant in any attempt to promote greater production and consumption of pulses. Enhancing these qualities in legumes thus offers affordable, scalable and enhanced health management to a very large number of undernourished children. The current phase II trial, if successful, will generate clinical and biological endpoints that will help inform the design and sample size of a future phase III trial with mortality as the primary endpoint.

**Study progress**
The modified feed was developed between August 2017 and March 2018. The final product has passed a full microbiological sterility testing following a pre-incubation at 30°C for 14 days, and an incubation on different culture media at 25°C, 37°C and 55°C for a further 7 days. The product is also confirmed to contain <40 µg/kg acrylamide, <400 HAU/g lectin, and <50 mg/kg tin, and was transported to Uganda in June 2018.

**Recruitment status**
The trial started enrolment on 6th July 2018 and will recruit over a period of 4–6 months. Primary data analysis and report-writing will take a further 4–6 months.

**Contact information**
The contact at Imperial College, for the MRC CIC grant scheme is confidenceinconcept@imperial.ac.uk. Contact for public enquiries should be made to Charles Bernard Okalebo, Mbale Clinical Research Institute, Pallisa Road, PO Box 1966, Mbale, Uganda (okabecha@gmail.com).

**Data availability**
All data underlying the results are available as part of the article and no additional source data are required.

**Competing interests**
No competing interests were disclosed.

**Grant information**
This study is funded by the MRC Confidence in Concept 2017 Scheme which is also supported by GCRF funding, and led by the Wellcome Trust Centre for Global Health Research (grant number 203077) in conjunction with the Institute for Global Health Innovation, at Imperial College London.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Acknowledgements**
We acknowledge the support and help of our colleagues in the nutrition unit and the Director and staff of the Mbale Regional Referral Hospital, in addition to Siraj Kijogo and Jennifer Adong.

**Supplementary material**
Supplementary File 1. The consent form to be signed/markd by the parent/guardian of each child.

Click here to access the data.

Supplementary File 2. Study information given to the parent/guardian of each child.

Click here to access the data.

Supplementary File 3. The complete set of case report forms used in the MIMBLE 2.0 study.

Click here to access the data.

**References**
5. Smith MI, Yatsunenko T, Manary MJ, et al.: Gut microbiomes of Malawian twin...
Open Peer Review

Current Peer Review Status: ✔️ ✔️

Version 1

Reviewer Report 24 September 2018

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The article describes very nicely the planned study. The study addresses a topic area that needs to be revised since the profile of children currently starting treatment as an inpatient has changed since the adoption of the community based-management of acute malnutrition (CMAM). If the area of the research, the CMAM programme has a well-functioning community component, children are likely to be admitted with less body function disturbance than when the current standard diets were developed.

In their introduction, Walsh and colleagues state that the standard formulations F75/F100 have not been able to reduce case-fatality and use this statement to partly justify the need for a new Formulation. Although case-fatality of complicated SAM remains high, there is evidence that the introduction of these standards formulations have been instrumental in obtaining the drop of case fatality from around 30 to 50% to below 10% provided that the adherence to the protocol is good. We think that this statement needs to be adjusted. However, it is correct that it has been difficult to reduce further the case-fatality of children requiring initial inpatient care and exploring the contribution of legumes floor and lactose free formula is important.

There are also areas of the methods section that may need to be revised to provide more details. Indeed more details are needed on the nutrition profile of the experimental formula, on the cooking process of the chickpea, on the rationale of using the energy density of 2kcal/ml and on the calculation of the sample size. Regarding details on the nutrition profile, it is important to have details on macro and micronutrients profile, amino-acid profile and PDCAAS, fatty acids profile and proportion of energy from omega-3 and from omega-6, phytic acid and iron and phytic acid and zinc ratios. This information will be instrumental in convincing the nutrition community that the formulation has a suitable nutrition profile. It will also help other researchers who may want to replicate the study.

Regarding the cooking process, it is important to provide details on how chickpea will be processed /cooked as we have recently learn that the tolerability of ready-to-use therapeutic food is not the same when the grains were roasted and when the floor was obtained by extrusion.
cooking. Inappropriate cooking may lead to poor tolerability with symptoms appearing after 10 to 14 days. Again, this information is critical for those who want to replicate the study.

Regarding the chosen energy density, the current recommendation is to initiate the nutrition rehabilitation with protein and energy intake that can be tolerated by the disturbed metabolic system. High intake of protein and energy at the initiation of treatment has been associated with increased case-fatality. Although for this research you have planned to restrict the intake of the experimental formula to the recommended energy intake of treatment phase 1, transition phase and phase 2, this may not be easy when the product will be used in routine. Thus, it is important to expand on the rational of the chosen density and its safety.

Regarding sample size, because the main benefits of the new formula is to improve gut function and reduce 90-days mortality, I could have expected that the sample size will be calculated using one or the two parameters. But because MUAC has been chosen as the better proxy of these 2 parameters, there is a need to define the expected difference or of the smallest clinically meaningful difference. The sample size should then be calculated based on this and taking into account that the planned study aims at demonstrating superiority of the experimental formula. A calculation, indicates that the sample size of 80 SAM per arm (those reaching the 90 days time-point) is adequate to demonstrate superiority if the difference between arms is 0.5 cm. Note that most regulatory bodies (including WHO) will not consider absence of difference in a superiority trial to be considered as proof of equivalence.

In addition to the additional details suggested above, I would like to suggest collection of some other parameters that may strengthen the evidence at acceptable extra-cost and efforts. Estimation of increment of lean mass difference may be added at a very limited cost using the simplest single-frequency bio-impedance analysis device. Indeed, plan based formulation are accused of promoting fat deposition rather than lean mass deposition. It will be important to demonstrate that this is not the case. Also, we recently published the results of study showing that reducing milk content of RUTF combined with the optimisation of phytic acid and iron molar ratio improves the correction of anaemia and iron deficiency. This finding suggests that some milk components inhibit iron absorption more than legumes anti-nutrients when iron and vitamin c contents of the diet are optimised. If similar results are observed with the experimental formula, the finding will provide an additional rationale for adopting this formula. Existing cheap laboratory tests will allow the addition of the assessment of haemoglobin and iron status at minimum increase in overall research budget.

**Is the rationale for, and objectives of, the study clearly described?**
Yes

**Is the study design appropriate for the research question?**
Yes

**Are sufficient details of the methods provided to allow replication by others?**
Partly

**Are the datasets clearly presented in a useable and accessible format?**
Not applicable
**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Undernutrition, therapeutic food formulations, child health, epidemiology and biostatistics

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

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Walsh and colleagues plan an open trial to compare the benefit of a legume flour-enriched formula with a conventional milk formula in Ugandan children with SAM. The objective is important and topical.

The proposed formula has two functions, to improve gut function while providing nutritional support. The improved formula is lactose-free, as brush border enzyme deficiency is common in SAM and undigested lactose will promote osmotic diarrhoea. However there are other components of a milk-based diet that may disrupt a damaged gut. For instance, there is increasing awareness of immune activation by b-casein sub-types. This has led to the discovery that some diagnoses of lactose intolerance in susceptible individuals may be due more correctly to immune activation by a1 b-casein. If this were widespread, then lactose-free skimmed milk powder may continue to disrupt small intestinal function through immune system activation. The study design has taken sufficient precaution to identify, record and act upon any adverse events, including any that may occur in response to the formula. The alternative may have been to develop a wholly plant-based formula. But this would have required much more preliminary work to prove it was palatable and capable of delivering adequate nutrition.

Grain legumes, in particular chickpea flour, has been chosen as the food to supplement lactose-free skimmed milk powder. This is the vector chosen to improve gut barrier function, by virtue of its fermentable dietary fibre content. Short chain fatty acids produced in the large bowel are pleiotropic agents with some of their benefit coming from improved barrier function. In terms of the RATIONALE of this proposal, it is not explained how a large bowel function can impact the small bowel, i.e., where nutrients are digested and absorbed. No evidence is presented for there being significant microbial fermentation in the terminal ileum allowing SCFA to act on local barrier function. Alternatively, SCFA action in the large bowel could have remote effects, possibly acting through L-cell released hormones, such as glucagon like peptides. This was not discussed.
However, the MIMBLE pilot study showed encouraging results.

The other role of the formula is to provide adequate nutrition. In the absence of this information, a crude calculation reveals approximately one third of the protein comes from the chickpea. The adequacy of the amino acid supply from the new formula should be calculated, at least in theory. This would match amino acid score and protein digestibility against the requirements. Methionine will be the first limiting amino acid in the legume protein, but as milk protein has an excellent score, the blend is likely to be adequate, but this should be demonstrated. Perhaps the greater issue is that grain legumes contain anti-nutritional compounds or factors (ANFs) that may reduce the bioavailability of dietary protein (and minerals) from the combined formula, not just the legume flour. Choice of variety, its processing and preparation can improve the situation but no details were given. Cooking will neutralise potential microbial hazard while improving digestibility. Cooking is also likely to reduce ANFs but excessive heat may promote Maillard and other reaction products that will reduce indispensable amino acid bioavailability. The METHODS should detail the legume variety, it should include a calculation of the digestibility-corrected amino acid score of the new formula and give an indication of the cooking/preparation procedure.

Markers of improved body composition would seem an appropriate choice for primary outcome. Considering the likely resources in the field, MUAC seems entirely appropriate. More accurate measures of Fat Free Mass gain are often too elaborate for use in a SAM setting. Shorter term measures of ‘protein status’ may be caught within the biochemistry and/or metabolomics screens.

**Is the rationale for, and objectives of, the study clearly described?**
Partly

**Is the study design appropriate for the research question?**
Yes

**Are sufficient details of the methods provided to allow replication by others?**
No

**Are the datasets clearly presented in a useable and accessible format?**
Not applicable

**Competing Interests:** No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.