Prevalences of inherited red blood cell disorders in pregnant women of different ethnicities living along the Thailand-Myanmar border [version 2; referees: 2 approved]

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Abstract

Background: Inherited red blood cell disorders are prevalent in populations living in malaria endemic areas; G6PD deficiency is associated with oxidant-induced haemolysis and abnormal haemoglobin variants may cause chronic anaemia. In pregnant women, microcytic anaemia caused by haemoglobinopathies mimics iron deficiency, complicating diagnosis and treatment. Anaemia during pregnancy is associated with morbidity and mortality. The aim of this study was to characterise the prevalence of G6PD deficiency and haemoglobinopathies among the pregnant population living along the Thailand-Myanmar border. Pregnant women attending antenatal clinics in this area belong to several distinct ethnic groups.

Methods: Data were available for 13,520 women attending antenatal care between July 2012 and September 2016. Screening for G6PD deficiency was done by fluorescent spot test routinely. G6PD genotyping and quantitative phenotyping by spectrophotometry were analysed in a subsample of women. Haemoglobin variants were diagnosed by HPLC or capillary electrophoresis and molecular methods. The prevalence and distribution of inherited red blood cell disorders was analysed with respect to ethnicity.

Results: G6PD deficiency was common, especially in the Sgaw Karen ethnic group, in whom the G6PD Mahidol variant allele frequency was 20.7%. Quantitative G6PD phenotyping showed that 60.5% of heterozygous women had an intermediate enzymatic activity between 30% and 70% of the population median. HbE, beta-thalassaemia trait and Hb Constant Spring were found overall in 15.6% of women. Only 45.2% of women with low percentage of HbA were carriers of mutations on the alpha globin genes.

Conclusions: Distribution of G6PD and haemoglobin variants varied among the different ethnic groups, but the prevalence was generally high throughout the cohort. These findings encourage the implementation of an extended program of information and genetic counselling to women of reproductive age.

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2

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program of information and genetic counselling to women of reproductive age and will help inform future studies and current clinical management of anaemia in the pregnant population in this region.

Keywords
G6PD deficiency, hemoglobinopathies, blood groups, ethnic groups, Thailand-Myanmar border, anemia, pregnancy

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Haemoglobinopathies are caused by a number of mutations on the genes that encode haemoglobin alpha and non-alpha chains (Weatherall, 2013). Alpha chains are encoded by four alpha genes located in pairs on the chromosome 16, while non-alpha chains are encoded by several genes on the beta-globin cluster on chromosome 11. Expression of non-alpha chains changes during embryonic, foetal and adult development; in children over one year of age and in adults, over 96% of haemoglobin is composed by two alpha and two beta chains(Hba) and between 2.2–3.5% by two alpha and two delta chains (HbA). Mutations that cause imbalance among the four chains are called thalassaemias (Hartveld & Higgs, 2010; Taher et al., 2017). Haemoglobinopathies are associated with a spectrum of reduced haemoglobin levels, ranging from very mild (ca 0.5g/dL, average reduction) to severe anaemia. The geographic distribution of abnormal haemoglobins corresponds to that of malaria before modern times, because these abnormalities confer some protection against malaria or its pathological effects (Haldane, 1949; Hutagalung et al., 1999; Siniscalco et al., 1961; Taylor & Fairhurst, 2014; Weatherall et al., 2010). The refugee and migrant populations living along the Thailand-Myanmar border, an area of low seasonal malaria transmission, are composed of several distinct ethnic groups: mainly Burman, Sgaw Karen and Poe Karen, followed by Mon, Kachin, Shan, and Rakhine. While a few studies have investigated the prevalence of haemoglobinopathies among Burman in central Myanmar (Than et al., 2005; Win et al., 2005), little is known about Karen and other ethnic minorities living in Karen state and along the Thailand border. There are scattered reports from Karen and Burman patients who have been resettled in high-income countries where the capacity to determine detailed genetic traits is possible (Lee, 2012; Phylipsen et al., 2010). G6PD deficiency has been studied in recent years in this border population (Bancone et al., 2014; Phompradit et al., 2011), but quantitative characterization of G6PD phenotypes in women has not been carried out yet at the population level.

The aim of this report is to describe the prevalence of IRD among the pregnant women attending routine antenatal screening at SMRU clinics along the Thailand-Myanmar border.

Methods

Study area and population

The Shoklo Malaria Research Unit is located in the north-western border of Thailand and has been providing free health assistance to the local refugee population for 30 years. The antenatal care (ANC) program was established for the early detection and treatment of malaria in 1986 for refugees and since 1998 for migrants. Women living in the catchment area of SMRU clinics are encouraged to attend the ANC as soon as they are aware of their pregnancy. Over 3,000 pregnant women of mainly Burman and Karen ethnicity register for ANC at SMRU clinics each year. SMRU’s free comprehensive antenatal, delivery and postpartum care has been described elsewhere (Hoogenboom et al., 2015). At the first consultation, demographic information is collected, an obstetric and medical history recorded, and detailed clinical examination done. Antenatal screening from July 2012 to September 2016 included examination of IRB in line with Thailand national guidelines. All women had language appropriate group counselling at this first visit about the different screening tests and this was followed by one on one
counselling with the option to opt out. Formal written consent was not required for the original routine blood sampling. Permission to waive consent from the individual patient for analysis of routine data was asked and obtained from the Ethic Committee of the Faculty of Tropical Medicine, Mahidol University (approval letter MUTM2017-041-01), and from Oxford University (OXTREC#583-16).

Definition of ethnicity
Pregnant women were asked to report the ethnicity of both parents as belonging to one of the following groups based on locally preferred terms for self-identification: Spaw Karen, Poe Karen, Burman, “Muslim”, Mon, Kachin, Pa Oh, Rakhine, Shan and “others”. The ethnicity of the woman was defined “mixed” when parents’ ethnicity differed. Of note, in this border area people of Islamic faith often self-identify as “Muslim” when asked about their ethnicity; the “Muslim” group is not an ethnic group, but rather a heterogeneous group of subjects with various ethnic backgrounds.

Laboratory analyses
In the central Haematology Laboratory, G6PD deficiency was screened by the NADPH Fluorescent Spot Test (FST, R&D Diagnostic, Greece). The FST allows detection of G6PD activity in a small volume of blood by supplying the substrates for the G6PD-mediated reduction of NADP+ to the naturally fluorescent NADPH. The fluorescence is detected in the mix spotted on paper by observation under long-wave UV light (ca 340nm) (Beutler & Mitchell, 1968). Although the FST can show varying degrees of fluorescent intensity, samples are classified either as deficient or as normal based on visual assessment. The threshold of enzymatic activity which produces a visually normal fluorescence has been estimated to be approximately 30% of normal (Bancone et al., 2015). As a result, samples classified as deficient include most of the severely mutated homozygote and hemizygote subjects; samples classified as normal include specimens from wild type subjects and from subjects with intermediate activity over the 30% threshold (e.g heterozygous women or individuals with milder mutations). G6PD quantitative phenotype was assessed by spectrophotometry carried out at 37°C according to the standard WHO protocol on whole blood depleted of WBCs (Beutler et al., 1977) in a subsample of sequentially recruited women. G6PD activity was calculated after normalization with Hb concentration and expressed as IU/gHb. The G6PD activity population median of 11.5 IU/gHb was established previously in the laboratory using the same technique on G6PD normal males (Bancone et al., 2014). In the same selected women, G6PD genotyping was performed using established PCR-RFLP protocols (Bancone et al., 2014). Genotyping for Mahidol was performed on all these women, while Chinese-4, Kaiping, Canton and Mediterranean variants were analysed only on women with either enzymatic activity below 80% of normal in Mahidol-wild type genotype or enzymatic activity below 30% and Mahidol-heterozygous genotype. The minimum size of the subsample analysed for G6PD quantitative phenotyping and genotyping was set at 300 subjects; this was calculated to allow at least 5 subjects in the smaller genotype group (homozygous mutated) based on the expected allele frequency of 15%.

Haemoglobinopathies
For haemoglobin typing, blood samples were analysed either by high-performance liquid chromatography (HPLC, using Biorad D-10™) at the local Mae Sot hospital or by Capillary Electrophoresis (CE, using a Capillaries II, Sebia, France) at the central SMRU Haematology Laboratory. Adult haemoglobin contains mainly HbA (2α2β, 96–98%) and HbA2 (2α2δ, 2.2–3.5%). Both HPLC and CE allow for diagnosis of beta-thalassaemia carriage (by mean of increased percentage of HbA2, usually over 5%), presumptive diagnosis of alpha-thalassaemia carriage (by mean of decreased percentage of HbA2, usually under 2.2%) and diagnosis of Hb structural variants such as HbE, HbC, HbS (by mean of appearance of retention peaks at specific elution times). Haemoglobin Constant Spring (HbCS), a non deletional alpha thalassaemia caused by a TAA>CAA mutation on the termination codon of alpha gene, was diagnosed by CE only, with a peak ≥20.5% in the HbC/SC retention zone. Common Asian deletion mutations on the alpha globin genes (3.7, 4.2, SEA) were analysed using a modified multiplex gap-PCR protocol (Chong et al., 2000). Molecular analysis was performed on an arbitrarily minimum sample size of 200 subjects with presumptive diagnosis of alpha-thalassaemia defined by low percentage of HbA2 (≤2.2% by HPLC or ≤2.1% by CE) in order to assess correlation between genotype and percentage of HbA2; an additional sample of at least 150 subjects with beta-thalassaemia trait was studied in order to assess prevalence of alpha-thalassaemia in this group and analyse variation of percentage of HbA2 in subjects with mutations on both alpha and non-alpha globin chains.

Statistical analysis
All women with available data in the timeframe of the study were included in the analysis. Data were entered and analysed in SPSS IBM SPSS Statistics for Windows, version 23.0 (IBM Corp., Armonk, N.Y., USA). Chi-square test was used for comparison of allelic frequencies among different ethnic groups. Normality of G6PD activity was tested by the Kolmogorov-Smirnov test.

Results
Study flow is summarized in Figure 1. A total of 15,512 pregnancies were registered at the ANC between July 2012 and September 2016. For women with more than one pregnancy during this time frame, successive pregnancies (n=1,754) were excluded from analysis. Complete demographic and ethnicity information were missing for 238 women, leaving a total of 13,520 analyzable women. G6PD FST screening and haemoglobin typing results were available in over 90% of women. Additional data on G6PD genotype and quantitative phenotype were analysed in a subsample of 317 sequential women who attended the clinics between August and September 2012. Between August 2012 and August 2013, alpha deletional mutations were analysed by PCR on 354 women presenting with a low percentage of HbA2 (≤2.2% by HPLC or ≤2.1% by CE) and on 159 women with beta-thalassaemia trait.

Prevalence and distribution of ethnic groups
A total of 12,605/13,520 (93.2%) women reported both parents to be of the same ethnicity and were therefore assigned to one of the nine common ethnic groups or to the pooled category of
uncommon ethnic groups called “others” (Table 1). Overall, the most represented ethnic group was the Sgaw Karen (44.3%), followed by Burman (28.5%), Poe Karen (12.4%) and “Muslim” (4.4%). The distribution of these four major ethnic groups reflected those of the general population attending the SMRU clinics with the Sgaw Karen comprising the vast majority of women attending in the refugee camp of MLA (71.3%) and Burman women representing 39.7% and 47.6% of the population attending the clinics for migrant population of WPA and MKT, respectively. Poe Karen were 9.1% to 16.9% in the three clinics, while “Muslim” were mainly in MLA camp for displaced persons (11.9%). The other ethnic groups were in very low numbers in all clinics.

**G6PD phenotype by fluorescent spot test**
A total of 12,427/13,520 (91.9%) women were screened for G6PD activity showing an overall prevalence of phenotypic deficiency of 2.9% (Table 2). Sgaw Karen and Mon ethnicities showed the highest prevalence of deficiency (over 4%), while all the other ethnic groups had less than 2% prevalence, with the “Muslim” and Rakhine at the lower end (close to 1%).

**G6PD genotypes according to ethnic group**
Among the subsample of 317 women, G6PD genotyping was performed for Mahidol, Canton, Kaiping, Chinese-4 and Mediterranean variants. Since the minor ethnic groups were hardly represented in this smaller sample, they were pooled together. The major mutation found was Mahidol, representing over 95% of all mutations in all ethnic groups, with the exception of Burmans in whom Kaiping, Canton and Chinese-4 were found globally at a polymorphic frequency (Table 3).

**Quantitation of G6PD activity by spectrophotometry**
The distribution of quantitative G6PD activity in 317 women is displayed in Figure 2, according to G6PD genotype (all mutations are pooled). G6PD activity in homozygous women for wild type allele was not normally distributed and had a median (IQR) of 11.76 (10.61–13.64) IU/gHb, similar to the median of 11.50 IU/gHb assessed previously in males; G6PD activity in the deficient homozygotes was not normally distributed and had a median (IQR) of 0.13 (0.11–0.25) IU/gHb. Both homozygous genotypes were therefore at the extremes of the activity spectrum, while heterozygous women had a wide distribution of activities from completely normal to “completely” deficient. The G6PD activity in 74 heterozygous women for the most prevalent Mahidol variant (Figure 3) showed a normal distribution with a mean (SD) of 7.38 (2.33) IU/gHb corresponding to 62.8% of the normal activity (based on the population median activity in females). According to this distribution, 6.6% of women had a G6PD activity in the range of deficiency (<30% normal activity), 60.5% were in the 30–70% activity range, and the remaining 32.9% were in the normal activity range (>70% normal activity).

**Haemoglobin variants**
Results of the haemoglobin typing by HPLC or Capillary Electrophoresis among 12,676 women are shown in Table 4. The highest HbE allelic frequency was found among women of Rakhine (23.2%) and Burman (11.0%) followed by the Mon ethnicities (7.7%). Sgaw Karen and Poe Karen had the lowest HbE allelic frequency among all the ethnic groups (1.0% and 1.7% respectively). Beta-thalassaemia had an allelic frequency lower than 5% in all ethnic groups, with a carrier prevalence less than 10%; the highest allelic frequency was found in Sgaw Karen.

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**Figure 1. Study flowchart.**
Table 1. Distribution of ethnic groups in the three Shoklo Malaria Research Unit clinics.

<table>
<thead>
<tr>
<th>Ethnicity</th>
<th>MKT</th>
<th>WPA</th>
<th>MLA</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sgaw Karen</td>
<td>1,081</td>
<td>1,470</td>
<td>3,443</td>
<td>5,994</td>
</tr>
<tr>
<td>Poe Karen</td>
<td>456</td>
<td>774</td>
<td>441</td>
<td>1,671</td>
</tr>
<tr>
<td>Burman</td>
<td>1,954</td>
<td>1,822</td>
<td>77</td>
<td>3,853</td>
</tr>
<tr>
<td>“Muslim”</td>
<td>15</td>
<td>11</td>
<td>573</td>
<td>599</td>
</tr>
<tr>
<td>Mon</td>
<td>72</td>
<td>56</td>
<td>14</td>
<td>142</td>
</tr>
<tr>
<td>Kachin</td>
<td>2</td>
<td>0</td>
<td>12</td>
<td>14</td>
</tr>
<tr>
<td>Pa Oh</td>
<td>46</td>
<td>501</td>
<td>10</td>
<td>106</td>
</tr>
<tr>
<td>Rakhine</td>
<td>55</td>
<td>14</td>
<td>1</td>
<td>70</td>
</tr>
<tr>
<td>Shan</td>
<td>9</td>
<td>10</td>
<td>6</td>
<td>25</td>
</tr>
<tr>
<td>“Mixed”</td>
<td>348</td>
<td>334</td>
<td>234</td>
<td>916</td>
</tr>
<tr>
<td>Other</td>
<td>66</td>
<td>48</td>
<td>16</td>
<td>130</td>
</tr>
<tr>
<td>Total</td>
<td>4,104</td>
<td>4,585</td>
<td>4,827</td>
<td>13,520</td>
</tr>
</tbody>
</table>

Table 2. G6PD phenotype by fluorescent spot test according to ethnic group.

<table>
<thead>
<tr>
<th>Ethnicity</th>
<th>Deficient</th>
<th>Normal</th>
<th>Total</th>
<th>% phenotypic deficiency</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sgaw Karen</td>
<td>226</td>
<td>5,110</td>
<td>5,336</td>
<td>4.2%</td>
</tr>
<tr>
<td>Poe Karen</td>
<td>25</td>
<td>1,525</td>
<td>1,550</td>
<td>1.6%</td>
</tr>
<tr>
<td>Burman</td>
<td>72</td>
<td>3,618</td>
<td>3,690</td>
<td>2.0%</td>
</tr>
<tr>
<td>“Muslim”</td>
<td>6</td>
<td>504</td>
<td>510</td>
<td>1.2%</td>
</tr>
<tr>
<td>Mon</td>
<td>7</td>
<td>128</td>
<td>135</td>
<td>5.2%</td>
</tr>
<tr>
<td>Kachin</td>
<td>0</td>
<td>12</td>
<td>12</td>
<td>0.0%</td>
</tr>
<tr>
<td>Pa Oh</td>
<td>1</td>
<td>99</td>
<td>100</td>
<td>1.0%</td>
</tr>
<tr>
<td>Rakhine</td>
<td>0</td>
<td>69</td>
<td>69</td>
<td>0.0%</td>
</tr>
<tr>
<td>Shan</td>
<td>1</td>
<td>23</td>
<td>24</td>
<td>4.2%</td>
</tr>
<tr>
<td>“Mixed”</td>
<td>20</td>
<td>860</td>
<td>880</td>
<td>2.3%</td>
</tr>
<tr>
<td>Other</td>
<td>2</td>
<td>119</td>
<td>121</td>
<td>1.7%</td>
</tr>
<tr>
<td>Total</td>
<td>360</td>
<td>12,067</td>
<td>12,427</td>
<td>2.9%</td>
</tr>
</tbody>
</table>

Table 3. G6PD genotypes among 317 women of different ethnicities.

<table>
<thead>
<tr>
<th></th>
<th>Wild type</th>
<th>Mahidol heterozygote</th>
<th>Chinese-4 heterozygote</th>
<th>Kaiping heterozygote</th>
<th>Mahidol homozygote</th>
<th>Mahidol-Canton</th>
<th>Total</th>
<th>Overall allelic frequency of all variants</th>
<th>Allelic frequency of Mahidol variant</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sgaw Karen</td>
<td>91</td>
<td>47</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>7</td>
<td>147</td>
<td>21.4%</td>
<td>20.7%</td>
</tr>
<tr>
<td>Poe Karen</td>
<td>34</td>
<td>12</td>
<td>0</td>
<td>1</td>
<td>4</td>
<td>0</td>
<td>51</td>
<td>20.6%</td>
<td>19.6%</td>
</tr>
<tr>
<td>Burman</td>
<td>67</td>
<td>9</td>
<td>1</td>
<td>2</td>
<td>0</td>
<td>1</td>
<td>80</td>
<td>8.8%</td>
<td>6.3%</td>
</tr>
<tr>
<td>“Muslim”</td>
<td>11</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>11</td>
<td>0.0%</td>
<td>0.0%</td>
</tr>
<tr>
<td>Other</td>
<td>6</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>7</td>
<td>7.1%</td>
<td>7.1%</td>
</tr>
<tr>
<td>“Mixed”</td>
<td>16</td>
<td>5</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>21</td>
<td>11.9%</td>
<td>11.9%</td>
</tr>
<tr>
<td>Total</td>
<td>225</td>
<td>74</td>
<td>2</td>
<td>4</td>
<td>11</td>
<td>1</td>
<td>317</td>
<td>16.3%</td>
<td>15.3%</td>
</tr>
</tbody>
</table>
Figure 2. G6PD activity (IU/gHb) assessed by spectrophotometer in 317 women.

Figure 3. Distribution of G6PD enzymatic activities among 74 Mahidol heterozygous women. Vertical red lines are the 30% and 70% of normal activity (based on the female population median, 100%).
Karen and Poe Karen (3.9%) and the lowest among women of the Rakhine and Burman ethnicities (0.9% and 0.7%, respectively). Haemoglobin Constant Spring (diagnosed only by capillary electrophoresis) was found in less than 2% of the women, with a higher prevalence among Burman (1.9%) and the lowest among Sgaw Karen (0.7%).

Mutations on the alpha globin chains
Among the 354 subjects with low HbA₂ and characterised by PCR (Table 5) less than half (45.2%) had mutations in the alpha genes, indicating that the percentage of HbA₂ alone was not a good indicator of alpha-thalassaemia carriage. The lowest HbA₂ percentage was associated with deletions on three alpha genes.

Table 4. Distribution of Hb variants diagnosed by HPLC or CE in the different ethnic groups.

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Sgaw Karen</th>
<th>Poe Karen</th>
<th>Burman</th>
<th>“Muslim”</th>
<th>Mon</th>
<th>Pa On</th>
<th>Rakhine</th>
<th>Shan</th>
<th>Kachin</th>
<th>Other</th>
<th>“Mixed”</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hb Normal</td>
<td>4,907</td>
<td>1,388</td>
<td>2,815</td>
<td>473</td>
<td>110</td>
<td>87</td>
<td>39</td>
<td>20</td>
<td>10</td>
<td>102</td>
<td>752</td>
<td>10,703</td>
</tr>
<tr>
<td>Beta-thalassaemia trait</td>
<td>421</td>
<td>123</td>
<td>60</td>
<td>17</td>
<td>4</td>
<td>4</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>3</td>
<td>49</td>
<td>683</td>
</tr>
<tr>
<td>Beta-thalassaemia trait with HbCS</td>
<td>3</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>4</td>
</tr>
<tr>
<td>Beta-thalassaemia/ HbE disease</td>
<td>1</td>
<td>1</td>
<td>3</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>9</td>
</tr>
<tr>
<td>HbEE</td>
<td>0</td>
<td>1</td>
<td>39</td>
<td>0</td>
<td>1</td>
<td>2</td>
<td>4</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td>49</td>
</tr>
<tr>
<td>HbE trait</td>
<td>103</td>
<td>51</td>
<td>726</td>
<td>30</td>
<td>19</td>
<td>9</td>
<td>24</td>
<td>2</td>
<td>1</td>
<td>20</td>
<td>71</td>
<td>1,056</td>
</tr>
<tr>
<td>HbE trait with HbCS</td>
<td>2</td>
<td>1</td>
<td>14</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>17</td>
</tr>
<tr>
<td>HbCS</td>
<td>34</td>
<td>17</td>
<td>58</td>
<td>7</td>
<td>2</td>
<td>2</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>8</td>
<td>130</td>
</tr>
<tr>
<td>Suspected HbH disease (HbA₂≤1.0)</td>
<td>2</td>
<td>2</td>
<td>6</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>10</td>
</tr>
<tr>
<td>Unknown Abnormal variant</td>
<td>6</td>
<td>1</td>
<td>6</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>15</td>
</tr>
<tr>
<td>Total</td>
<td>5,479</td>
<td>1,586</td>
<td>3,727</td>
<td>528</td>
<td>136</td>
<td>104</td>
<td>69</td>
<td>24</td>
<td>13</td>
<td>126</td>
<td>884</td>
<td>12,676</td>
</tr>
<tr>
<td>HbE carrier prevalence</td>
<td>1.9%</td>
<td>3.4%</td>
<td>21.0%</td>
<td>5.9%</td>
<td>14.7%</td>
<td>10.6%</td>
<td>42.0%</td>
<td>12.5%</td>
<td>7.7%</td>
<td>15.9%</td>
<td>8.4%</td>
<td>8.9%</td>
</tr>
<tr>
<td>HbE allelic frequency</td>
<td>1.0%</td>
<td>1.7%</td>
<td>11.0%</td>
<td>2.8%</td>
<td>7.7%</td>
<td>6.3%</td>
<td>23.2%</td>
<td>4.2%</td>
<td>3.8%</td>
<td>7.9%</td>
<td>4.2%</td>
<td>4.6%</td>
</tr>
<tr>
<td>Beta-thalassaemia carrier prevalence</td>
<td>7.8%</td>
<td>7.9%</td>
<td>1.7%</td>
<td>3.4%</td>
<td>2.9%</td>
<td>3.8%</td>
<td>1.4%</td>
<td>8.3%</td>
<td>7.7%</td>
<td>2.4%</td>
<td>5.7%</td>
<td>5.5%</td>
</tr>
<tr>
<td>Beta-thalassaemia allelic frequency</td>
<td>3.9%</td>
<td>3.9%</td>
<td>0.8%</td>
<td>1.7%</td>
<td>1.5%</td>
<td>1.9%</td>
<td>0.7%</td>
<td>4.2%</td>
<td>3.8%</td>
<td>1.2%</td>
<td>2.8%</td>
<td>2.7%</td>
</tr>
<tr>
<td>HbCS carrier frequency</td>
<td>0.7%</td>
<td>1.2%</td>
<td>1.9%</td>
<td>1.3%</td>
<td>1.5%</td>
<td>1.9%</td>
<td>0.0%</td>
<td>0.0%</td>
<td>7.7%</td>
<td>0.8%</td>
<td>0.9%</td>
<td>1.2%</td>
</tr>
</tbody>
</table>

Table 5. Median HbA₂ percentage according to alpha chain genotype in subjects with presumptive alpha-thalassaemia or beta-thalassaemia trait.

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Presumptive alpha-thalassaemia (HbA₂≤2.2)</th>
<th>Median (IQR) %HbA₂</th>
<th>Beta-thalassaemia trait</th>
<th>Median (IQR) %HbA₂</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>194</td>
<td>1.90 (1.80-2.10)</td>
<td>119</td>
<td>5.20 (4.70-5.70)</td>
</tr>
<tr>
<td>αα/α-3.7</td>
<td>124</td>
<td>1.90 (1.80-2.00)</td>
<td>37</td>
<td>5.20 (4.60-5.80)</td>
</tr>
<tr>
<td>αα/α -4.2</td>
<td>1</td>
<td>1.70</td>
<td></td>
<td></td>
</tr>
<tr>
<td>αα/-SEA</td>
<td>8</td>
<td>1.75 (1.63-2.12)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>α-3.7/α-3.7</td>
<td>20</td>
<td>1.85 (1.80-1.90)</td>
<td>3</td>
<td>6.60</td>
</tr>
<tr>
<td>α-3.7/-SEA (HbH)</td>
<td>7</td>
<td>0.60 (0.40-1.00)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
(HbH disease), (P<0.01 vs all others), while subjects with two or one mutation had increasingly higher HbA2 percentage, but not significantly different from those without mutations (Table 5). Since this was a selected group of women, the prevalence and distribution of mutations analysed (3.7, 4.2 and SEA) cannot be considered representative of the general pregnant population. Among the consecutively enrolled 159 beta-thalassaemia carriers the prevalence of concurrent alpha-thalassaemia carriage was 25.3% and in this group there was no difference in percentage of HbA2 between subjects with alpha deletions and those without. Overall, the most common deletion was -3.7 followed by SEA.

**Discussion**

In Myanmar, the population is divided officially into eight main ethnicities, which the government has further classified into 135 different indigenous ethnic groups. The majority group Burman make up 68% of the country’s population of 55 million, with the Shan (9%), Karen (7%), Rakhine (or Arakanese) (4%) and Mon (2%) comprising the largest minority groups. In this area of the Thailand-Myanmar border, the Karen group predominates and this is further classified into Sgaw and Poe. Patients of Sgaw Karen, Burman, Poe Karen and “Muslim” ethnicity represent the four local major ethnic groups who seek health care at border SMRU clinics.

G6PD deficiency is relatively common in all ethnic groups with phenotypic deficiency prevalence in women ranging from 1% of most groups (corresponding to the observed allelic frequency of 10%) to a maximum of over 4% (corresponding to the observed allelic frequency of 20%) in the Sgaw Karen. These data confirm previous observations in the male population on the Thailand-Myanmar border (Bancone et al., 2014; Louicharoen et al., 2009; Phompradit et al., 2011), inside Myanmar (Arnolda et al., 2015; Matsuoka et al., 2004; Nuchprayoon et al., 2008) and in Kachin state adjacent to the Myanmar-China border (Li et al., 2015). Current G6PD qualitative field tests can only detect marked deficiency, and women classified as “G6PD normal” are a rather heterogeneous group of subjects with G6PD activity ranging from 30% to 100% of normal, whose haemolytic risk when treated with oxidative drugs varies widely. Characterization of quantitative G6PD phenotype in a large population of females has never been carried out before and represents a resource for informing treatment for several infectious diseases in women living in an endemic area. Nitrofurantoin, a known precipitant of haemolysis in G6PD deficient patients (Youngster et al., 2010), is recommended as first line treatment for urinary tract infections in pregnancy (Gupta et al., 2011), due to its safety, low cost, and efficacy. Primaquine cannot be prescribed in pregnant women, but it is indicated postpartum for women infected with *P. vivax* during pregnancy and is a cornerstone of vivax malaria elimination. Our data show that over 60% of heterozygote women have a G6PD enzymatic activity over the threshold of deficiency for field tests but in the intermediate range (i.e. lower than the normal), and so could be at risk of drug induced haemolysis with certain primaquine regimens (Chu et al., 2017). Furthermore, the slightly skewed distribution of G6PD activities among heterozygous women might suggest some somatic selection against deficient RBCs as seen in the severe variant Mediterranean (Rinaldi et al., 1976); more investigations will be needed to establish whether this is specific to pregnancy or to the Mahidol variant.

Haemoglobin variants are common in this population, affecting at least 15% (HbE and beta-thalassaemia) and possibly up to more than 40% (including alpha-thalassaemia) of women overall. There are marked differences in the prevalence of HbE and beta-thalassaemia in different ethnic groups (as described previously (Win et al., 2005)) and this peculiar distribution, combined with a low rate of intermarriage between ethnic groups, suggests that the population may have partial protection from the deleterious beta-thalassaemia/HbE syndrome. In fact only 9 women in the entire cohort (0.07%) were found to have beta-thalassaemia/HbE syndrome, significantly below what would be expected in a random mating population (χ² =23.1, P<0.001). Furthermore, within the Sgaw Karen ethnicity only, the observed frequency of beta-thalassaemia/HbE genotype was lower than expected (χ² =9.14, P<0.05) suggesting a further associated reduced survival to reproductive age. Molecular characterization of alpha-thalassaemia was only performed in subjects with a presumptive diagnosis by HPLC or CE and in a subsample of beta-thalassaemia carriers. Data presented here confirm that, with the exclusion of HbH disease, a percentage of HbA2 ≤2.2 was not specific for alpha-thalassaemia trait, as over half of the women identified by this criteria had a normal genotype (Van Delft et al., 2009). The assessed higher prevalence of alpha-thalassaemia among the beta-thalassaemia carriers (25.3%) seems to be closer to the true prevalence in the population and it is higher than that estimated only by the low percentage of HbA2 (16.9%).

While the main reasons for investigating G6PD deficiency and Hb variants in pregnant women is the clinical management of anaemia and treatment with antimicrobial agents, the results carry implications for the offspring of the women tested. Communicating this type of information to women in this population with low health literacy in meaningful language is challenging. When G6PD deficiency is diagnosed, the staff counsel the woman about which drugs and food should be avoided and give her a card explaining the diagnosis and contraindicated medications. When beta-thalassaemia and HbE are diagnosed, the woman is informed that she might experience weakness or other anaemia symptoms during the pregnancy and a card with the diagnosis and a short description is given to the patient. During this simple counselling the woman is informed she might have passed her genetic trait to her present foetus and might pass it in future pregnancies; the possibility that the offspring might inherit the abnormal Hb trait from both parents and be severely affected represents an important, but difficult to convey, part of the counselling. The challenge of counselling about often asymptomatic diseases with complex implications is significant (McGready et al., 2014) and could result in dire unintended consequences in some individuals. In countries where haemoglobinopathies are common, several screening approaches have been developed (for
a review see Amato et al., 2014). Cost/benefit studies have shown that prevention programs are highly cost effective (Koren et al., 2014) and a means to reduce suffering for patients and families (Ballantyne et al., 2009). In Myanmar, there is no routine practice of prenatal screening for Hb variants, and this kind of testing would only be available to private patients consulting at specialty clinics in major urban centres. In Thailand, the screening is performed by HPLC on voluntary basis at the first ANC appointment. When the mother is found to be a carrier, the father of the baby is also tested and if found to be carrier, molecular investigations are performed. In the low-resource setting of SMRU, Hb testing for the mother alone costs approximately the same amount as all other investigations performed at the first ANC consultation combined. Hb testing for the father is not routinely performed due to cost, and pre-natal foetal diagnosis is not possible. Furthermore, SMRU is not equipped to offer long-term clinical care for subjects with transfusion-dependent thalassaemia, and the costs for refugees or migrants would be a limiting factor even when referral to a reference centre in Thailand is possible. Safe termination of pregnancy is not accessible to most migrant and refugee women in this border area and unsafe abortions are a cause of maternal morbidity and mortality in these vulnerable communities (Turner et al., 2012). Despite these challenges and limitations, the current data on the high prevalence of HbE, alpha and beta-thalassaemia warrant a continuation of screening and encourage the implementation of a more extended program of early information and counselling to girls and women of reproductive age among the population. The pioneering and remarkable example of the thalassaemia screening carried out by the “Centro Studi Microcitemie Roma” during 37 years among young students in Latium, Italy, (Amato et al., 2014) shows that information, screening of relatives, and counselling are major factors in the success of programs for the prevention of severe haemoglobinopathies. For long term sustainability, laboratory testing can be carried out with cheaper techniques, such as single-tube osmotic fragility test (OFT), dichlorophenolindophenol (DCIP) for HbE, and microscopic examination following a step-wise approach whereby the most expensive tests are only performed on samples that result abnormal by initial screening. This approach, especially if offered as part of a strong preconception healthcare package, including partner testing where appropriate, genetic counselling, folic acid supplementation, and effective and acceptable family planning provision, could substantially decrease the suffering of vulnerable families. Such a program would require community engagement and human resource development to equip local staff with the specialized skills of sensitive and responsive genetic counselling for individuals with limited background science knowledge.

In conclusion, the current data show that G6PD deficiency and abnormal haemoglobins are common among pregnant women in this area of the Thailand-Myanmar border, and likely contribute to increase the proportion of women with anaemia. In women with congenital Hb variants, anaemia can be present before pregnancy and might worsen during gestation, and requirement for iron supplementation can be difficult to assess. The current population data will help inform ongoing efforts to optimize the clinical management of anaemia in the local pregnant population by investigating newer marker of iron deficiency anaemia (for example, hepcidin), which might be more reliable in such context as compared to classic markers (Bah et al., 2017; Wray et al., 2017).

Data availability
Due to ethical and security considerations, the data that supports the findings in this study can be accessed only through the Data Access Committee at Mahidol Oxford Tropical Medicine Research Unit (MORU). The data sharing policy can be found here: http://www.tropmedres.ac/data-sharing. The application form for datasets under the custodianship of MORU Tropical Network can be found in Supplementary File 1.

Ethical statement
Ethical approval for the study was granted by the Faculty of Tropical Medicine, Mahidol University (MUTM 2017-041-01) and by Oxford University (OXTREC#583-16).

Competing interests
No competing interests were disclosed.

Grant information
This work was supported by the Wellcome Trust [106698], Major Overseas Programme–Thailand Unit, which supports the Shoklo Malaria Research Unit, part of the Mahidol Oxford University Research Unit; [089179] to GB; and 5% Initiative of French Government [12IN1211] for analyses of Hb typing.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgments
The authors would like to thank all the women participating in the cohort. They also thank all the staff of SMRU clinics and laboratories for their hard work and dedication. In particular they wish to thank Raweewan Somsakchaicharon, Alisara Khamrsi, Lalita Poodpanya and Paw Khu Moo from the central Haematology Laboratory.

Supplementary material
Supplementary File 1: Application form for datasets under the custodianship of Mahidol Oxford Tropical Medicine Research Unit (MORU) Tropical Network.

Click here to access the data.
References


Open Peer Review

Current Referee Status: ✔️ ✔️

Version 2

Referee Report 13 November 2017
doi: 10.21956/wellcomeopenres.14103.r27543

Issarang Nuchprayoon
Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand

I have read the revised version and all points were addressed. The responses and reasons were reasonable. Well done!

Competing Interests: No competing interests were disclosed.

Referee Expertise: molecular genetics, G6PD, blood groups, clinical care of thalassemia

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 1

Referee Report 25 September 2017
doi: 10.21956/wellcomeopenres.13359.r25455

Thomas N. Williams 1,2
1 Centre for Geographic Medicine Research-Coast, Kenyan Medical Research Institute (KEMRI)-Wellcome Trust Research Programme, Kilifi, Kenya
2 Imperial College, London, UK

Thanks for these changes. I think the manuscript is now acceptable. Well done!

Competing Interests: No competing interests were disclosed.

Referee Expertise: Haemoglobinopathies, epidemiology, clinical research, malaria, infectious diseases

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
This study is a valuable study based on its large sample size of pregnant women of rarely studied ethnic group, namely Sgaw Karen in comparison with Burman.

The subject of study; inherited red cell variants are valid in itself. It is controversial whether heterozygous G6PD mutation, thalassemia traits, and ABO, Rh blood groups are 'disorders'. People with all of these conditions do not have anemia in their usual state. Only people with compound heterozygous thalassemia 'diseases', homozygous HbE (HbEE), and HbH will have some degree of anemia. The causes of anemia in pregnant women are by far commonly caused by iron deficiency, which is not addressed in this study. Therefore the reference to anemia in pregnancy should be de-emphaized in the rationale and conclusion of this study.

G6PD results in this report does confirm other studies in Myanmar ethnic groups, many of which were reported in males. One additional citation that the author should consider is a report from Louicharen et al, 2009, who studied prevalence of G6PD mutations in Karen of suan peung, at a Thai-Myanmar border. In this paper in which details are reported in supplementary article in Science website. the prevalence of G6PD deficiency 20%, with predominantly (94%) Mahidol variant.

The methodology for estimating prevalence of alpha-thalassemia trait in this paper is controversial. Using low HbA2 is known to be unreliable and in fact the authors showed only 44.1% specificity. Since it is known how much is alpha-tha are missed in people with normal A2, the total prevalence is therefore not known for certainty. The prevalence of alpha-thalassemia in subset of studies in known beta-thal trait population may be more accurate estimation assuming all subjects were tested, but need to be stratified by each ethnic group. The author may wish state a conservative estimate in each ethnic group, like at least 25%, instead of a specific proportion.

Due to large number of Karen and Burmans in this study, the author may wish to point out the value of this paper in population genetics. Their difference in prevalence was a founder effect and due to estimation of G6PD Mahidol variant emerging around 1500 years ago as protective factor against malaria, the Burman may be a more recent population in this region.

In the final conclusion paragraph, the author should made a summary statement of their key findings on prevalence of variants but not about anemia since this paper did not address the issue. A mention of newer marker for iron deficiency detection is irrelevant and not useful in this resource-limited region. Suggestion of counseling is irrelevant and impractical here.

References

Is the work clearly and accurately presented and does it cite the current literature?
Partly

Is the study design appropriate and is the work technically sound?
Partly

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
Yes

Are all the source data underlying the results available to ensure full reproducibility?
Yes

Are the conclusions drawn adequately supported by the results?
Yes

**Competing Interests:** No competing interests were disclosed.

**Referee Expertise:** molecular genetics, G6PD, blood groups, clinical care of thalassemia

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

---

**Author Response 23 Oct 2017**

**Germana Bancone,** Shoklo Malaria Research Unit, Thailand

We thank the reviewer for his useful comments and suggestions. Below the answer to his questions.

**Q1)** The subject of study; inherited red cell variants are valid in itself. It is controversial whether heterozygous G6PD mutation, thalassemia traits, and ABO, Rh blood groups are ‘disorders’. People with all of these conditions do not have anemia in their usual state. Only people with compound heterozygous thalassemia ‘diseases’, homozygous HbE (HbEE), and HbH will have some degree of anemia. The causes of anemia in pregnant women are by far commonly caused by iron deficiency, which is not addressed in this study. Therefore the reference to anemia in pregnancy should be de-emphaized in the rationale and conclusion of this study.

**A1)** Anaemia is the most common medical problem of the pregnant population under study and is associated with maternal mortality (McGready, Boel et al. 2012) but a systematic analysis of genetic causes of anemia has never been undertaken. We think that, although analyzing these markers is interesting in itself, we cannot avoid highlighting that our primary interest is their contribution to anaemia during pregnancy; our plan is also to analyse soon clinical data in relation to the findings presented in this manuscript. Following the reviewers suggestions, we have re-phrased some of the text in the introduction and conclusion to clarify our message.

Q2) G6PD results in this report does confirm other studies in Myanmar ethnic groups, many of which were reported in males. One additional citation that the author should consider is a report
from Louicharen et al, 2009\textsuperscript{1}, who studied prevalence of G6PD mutations in Karen of suan peung, at a Thai-Myanmar border. In this paper in which details are reported in supplementary article in Science website. the prevalence of G6PD deficiency 20%, with predominantly (94%) Mahidol variant.

A2) We have added the suggested citation

Q3) The methodology for estimating prevalence of alpha-thalassemia trait in this paper is controversial. Using low HbA2 is known to be unreliable and in fact the authors showed only 44.1\% specificity. Since it is known how much is alpha-tha are missed in people with normal A2, the total prevalence is therefore not known for certainty. The prevalence of alpha-thalassemia in subset of studies in known beta-thal trait population may be more accurate estimation assuming all subjects were tested, but need to be stratified by each ethnic group. The author may wish state a conservative estimate in each ethnic group, like at least 25\%, instead of a specific proportion.

A3) As we answered also to the other reviewer, we have modified all the text and Tables to improve accuracy and clarity on this point.

Q4) Due to large number of Karen and Burmans in this study, the author may wish to point out the value of this paper in population genetics. Their difference in prevalence was a founder effect and due to estimation of G6PD Mahidol variant emerging around 1500 years ago\textsuperscript{1} as protective factor against malaria, the Burman may be a more recent population in this region.

A4) Although this is an interesting point to consider, the current data were not collected or analyzed to support or contradict evolutionary dynamics of Karen and Burman populations, therefore we prefer not to address the topic.

Q5) In the final conclusion paragraph, the author should made a summary statement of their key findings on prevalence of variants but not about anemia since this paper did not address the issue. A mention of newer marker for iron deficiency detection is irrelevant and not useful in this resource-limited region. Suggestion of counseling is irrelevant and impractical here.

A5) We have revised the text of the conclusive paragraph. We strongly believe that, having found a pretty high prevalence of abnormal Hb variants, a discussion on counselling pregnant women is warranted in this setting. Though this counselling is time-intensive and challenging, it is feasible in low resource settings with appropriately trained and sensitive local staff. Furthermore, better diagnostic tools for iron-deficient anaemia during pregnancy have been shown to be particularly useful in populations with a high prevalence of haemoglobin disorders (Wray, Allen et al. 2017); our data bring evidence on the widespread distribution of IRD and on the need for testing in resource-limited areas.

\textbf{Competing Interests:} I have no competing interests

Referee Report 04 September 2017

doi:10.21956/wellcomeopenres.13359.r25454

\textbf{Thomas N. Williams} \textsuperscript{1,2}

\textsuperscript{1} Centre for Geographic Medicine Research-Coast, Kenyan Medical Research Institute
This is a nice summary of an impressive study of red cell disorders in more than 13,000 pregnant women by the team at the Shoklo Malaria Research Unit on the Thai-Myanmar border.

**Major comments**

1) The background to the study is rationalized on the basis of the potential importance of inherited red cell disorders as causes and confounders of anaemia in pregnant women within the region. The inclusion of data on ABO and Rhesus in this paper, though interesting, does not fit well within this rational framework. Neither are particularly relevant to the main story. I would either drop those data or alter the rationale to justify their inclusion.

2) The referencing could be improved throughout the introduction. For example, a reference for Lyonisation and G6PD activity could be added, and the third paragraph, which contains a lot of technical information, should be referenced along the way rather than through the bunching of 4 (slightly random) references at the end.

3) Introduction paragraph 3 could be tightened up. The first sentence is a bit confused: “..structural variants and deletional mutations on the hemoglobin …. chains” is a bit of a mixture of nomenclature more usually specific to either the protein or the gene. The “deletional mutations” more commonly refer to the gene than the protein and probably more correctly should be referred to as “genetic deletions” rather than mutations (which more commonly relate to sequence changes than deletions). Further down, “…97% of hemoglobin is composed of two alpha and two beta chains” – the authors should clarify that they’re talking about ADULT haemoglobin (HbA). The references on the geographic distribution with malaria are a bit random and refer to different IRDs. A recent authoritative review would probably be a better way to go.

4) Methods: Lab analyses. It would be very helpful if the authors explained here on behalf of the non-expert what the G6PD spot test is designed to do and at what level of normal G6PD activity the test turns positive. A major issue with this test, as the authors acknowledge, is that it is a binary test that shows sufficient / deficient only. It is not designed to pick up heterozygotes and is only partially sensitive for homozygotes, depending on the genotype involved and the degree to which it perturbs G6PD production. Some discussion of this in the methods section would help the non-expert reader.

5) In the same section, the authors state that the quantitative test was conducted in “selected women”. It took me some digging to find that it was actually conducted in “a sub-set of sequentially recruited [and therefore “unselected”] women”. This could be better phrased at this point.

6) It would help the reader if the flow chart (Figure 1) could be amended to make the above more clear with regard to the quantitative test and the genotyping for both G6PD and other conditions.

7) Methods; haemoglobinopathies. The authors used an HbA2 of <2.2 or <2.1% (depending on the method) to define “presumptive” carrier status for alpha-thalassaemia. As the authors state later in the paper, this is neither a sensitive nor specific approach to the diagnosis of the alpha-thalassaemias overall. It's almost useless for the carrier forms of the common minor single gene -3.7 and -4.2kb deletional forms of the condition. As such, the authors’ focus on this sub-group for further work up for alpha thalassaemias constitutes a biased sample. It would really have been better from the perspective of trying to determine the prevalence of these conditions if the authors had centred their genetic testing on an unselected sample of women. This is the approach they took for G6PD genotyping which therefore produced figures that were more understandable. The figures given for the prevalence of the various alpha thalassaemias are difficult to interpret and probably very misleading because of the way in which the patients were sampled.

8) For this reason, I would remove the rows regarding “suspected alpha thalassaemia” from table 5 – the authors say later on that half of the sub-sample they tested were normal genetically, so this row will mislead the casual reader. HbCS and HbE are more legitimate as these have their own HPLC peaks.
9) Discussion, paragraph 2 seems a bit odd and comes rather out of the blue. This is the first mention of anthropology that I can find in the paper and the use of these IRDs as anthropological tools is put up and shot down in a single sentence!

10) Authors could be more clear that their estimate of G6PDd on the basis of their spot test result will be an underestimate, since homozygosity for less severe variants will not always be picked up (as shown by the outlier on Fig 2).

11) Discussion page 10 paragraph 2 left column. The authors say that all women with “Hb variants” are informed of the results and counseled that they may experience weakness etc in pregnancy. I assume that this is not the case for “suspected alpha thalassaemia”? This would give rise to a great deal of unnecessary anxiety if half don’t actually have the condition anyway and in the case of the -3.7 and -4.2 deletions they are clinically silent when inherited alone. The whole area of how to approach antenatal counseling in populations with a high prevalence of multiple conditions is highly complex and the discussion of this element could be tightened up a bit.

Minor
1) Though it’s an editorial decision, I would have assumed that since this is a UK based journal the spelling should be in UK English throughout.
2) Abstract Methods: Blood groups were “diagnosed” makes it sound as if these are a disease! I would change to “determined” or similar.
3) “resource rich countries” – preferred current term is “high-income countries”
4) Final paragraph of the introduction could probably be condensed and moved to join the methods section. I would replace by a short sentence summarizing the aims of the current study.
5) Please give the make and model of the HPLC machine used for the haemoglobin phenotyping.

Could do with a good proof-read throughout. A few examples I picked up:
1) Introduction line 3 “…living in area of past…” should read “…living in areas of past…”
2) Abstract Methods: Data were available rather than data was available (data are plural in this context).

3) page 4 line one column 2 – “SAE” should read “SEA”.
4) Table 6 – use the Greek symbol for alpha rather than the letter A.
5) page paragraph 2 left column “found to be carrier..” should read “found to be carriers..”

References

Is the work clearly and accurately presented and does it cite the current literature?
Partly

Is the study design appropriate and is the work technically sound?
Partly

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
Partly

Are all the source data underlying the results available to ensure full reproducibility?
No

Are the conclusions drawn adequately supported by the results?
Partly

**Competing Interests:** No competing interests were disclosed.

**Referee Expertise:** Haemoglobinopathies, epidemiology, clinical research, malaria, infectious diseases

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

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Author Response 23 Oct 2017

**Germana Bancone**, Shoklo Malaria Research Unit, Thailand

We thank the reviewer for his useful comments and suggestions. Below the answers to his questions.

**Major comments**

Q1) The background to the study is rationalized on the basis of the potential importance of inherited red cell disorders as causes and confounders of anaemia in pregnant women within the region. The inclusion of data on ABO and Rhesus in this paper, though interesting, does not fit well within this rational framework. Neither are particularly relevant to the main story. I would either drop those data or alter the rationale to justify their inclusion.

A1) Although the description of ABO and Rhesus blood groups was meant to add information on the genetic background of ethnic groups, we agree with the reviewer that it does not fit well with the framework of the manuscript. We have therefore decided to drop this part.

Q2) The referencing could be improved throughout the introduction. For example, a reference for Lyonisation and G6PD activity could be added, and the third paragraph, which contains a lot of technical information, should be referenced along the way rather than through the bunching of 4 (slightly random) references at the end.

A2) We have added more references in the introduction. The 4 references mentioned by the reviewer only concerned the selection by malaria of haemoglobinopathies.

Q3) Introduction paragraph 3 could be tightened up. The first sentence is a bit confused: “..structural variants and deletional mutations on the hemoglobin …. chains” is a bit of a mixture of nomenclature more usually specific to either the protein or the gene. The “deletional mutations” more commonly refer to the gene than the protein and probably more correctly should be referred to as “genetic deletions” rather than mutations (which more commonly relate to sequence changes than deletions). Further down, “…97% of hemoglobin is composed of two alpha and two beta chains” – the authors should clarify that they’re talking about ADULT haemoglobin (HbA). The references on the geographic distribution with malaria are a bit random and refer to different IRDs. A recent authoritative review would probably be a better way to go.

A3) We have reviewed and corrected the paragraph to include the reviewer’s suggestions.
Q4) Methods: Lab analyses. It would be very helpful if the authors explained here on behalf of the non-expert what the G6PD spot test is designed to do and at what level of normal G6PD activity the test turns positive. A major issue with this test, as the authors acknowledge, is that it is a binary test that shows sufficient / deficient only. It is not designed to pick up heterozygotes and is only partially sensitive for homozygotes, depending on the genotype involved and the degree to which it perturbs G6PD production. Some discussion of this in the methods section would help the non-expert reader.

A4) We have revised the text to better explain the G6PD fluorescent spot test.

Q5) In the same section, the authors state that the quantitative test was conducted in “selected women”. It took me some digging to find that it was actually conducted in “a sub-set of sequentially recruited [and therefore “unselected”] women”. This could be better phrased at this point.

A5) We agree with the reviewer that this part of the method was not phrased clearly and we have modified it.

Q6) It would help the reader if the flow chart (Figure 1) could be amended to make the above more clear with regard to the quantitative test and the genotyping for both G6PD and other conditions.

A6) The Flow chart has been modified to exclude results of ABO and Rh blood groups and to clarify G6PD and Hb genotyping.

Q7) Methods; haemoglobinopathies. The authors used an HbA2 of <2.2 or <2.1% (depending on the method) to define “presumptive” carrier status for alpha-thalassaemia. As the authors state later in the paper, this is neither a sensitive nor specific approach to the diagnosis of the alpha-thalassaemias overall. It’s almost useless for the carrier forms of the common minor single gene -3.7 and -4.2kb deletional forms of the condition. As such, the authors’ focus on this sub-group for further work up for alpha thalassaemias constitutes a biased sample. It would really have been better from the perspective of trying to determine the prevalence of these conditions if the authors had centred their genetic testing on an unselected sample of women. This is the approach they took for G6PD genotyping which therefore produced figures that were more understandable. The figures given for the prevalence of the various alpha thalassaemias are difficult to interpret and probably very misleading because of the way in which the patients were sampled.

A7) We thank the reviewer for pointing out this issue that we got wrong in the first version of the manuscript. At the time we started the screening we were not fully aware of the low specificity of the Hb typing for alpha-thalassaemia and one of our aims was to assess the diagnostic power of CE/HPLC for all hemoglobin variants. Nevertheless we agree that the way we presented the data was not appropriate and that the prevalence of alpha gene mutations in the subgroup of women with low HbA2 is not representative of the general population. We have therefore modified the text and the Table. This has also allowed correcting the numbers of women with low HbA2 tested by PCR which is 354 and not 374 as reported previously.

Q8) For this reason, I would remove the rows regarding “suspected alpha thalassaemia” from table 5 – the authors say later on that half of the sub-sample they tested were normal genetically, so this row will mislead the casual reader. HbCS and HbE are more legitimate as these have their own HPLC peaks.

A8) This has been done.

Q9) Discussion, paragraph 2 seems a bit odd and comes rather out of the blue. This is the first mention of anthropology that I can find in the paper and the use of these IRDs as anthropological
tools is put up and shot down in a single sentence!
A9) Since we have compared all these genetic markers between ethnic groups, one might argue that they could be used to estimate genetic differences between the ethnicities. We have removed this sentence together with the analysis of ABO/Rh blood groups.

Q10) Authors could be more clear that their estimate of G6PDd on the basis of their spot test result will be an underestimate, since homozygosity for less severe variants will not always be picked up (as shown by the outlier on Fig 2).
A10) We have added and clarified a few sentences to emphasize this point which is already well described in the literatures. With this analysis we wanted to concentrate the attention on the distribution of G6PD activities among heterozygous women which has not been studied extensively before.

Q11) Discussion page 10 paragraph 2 left column. The authors say that all women with “Hb variants” are informed of the results and counseled that they may experience weakness etc in pregnancy. I assume that this is not the case for “suspected alpha thalassaemia”? This would give rise to a great deal of unnecessary anxiety if half don’t actually have the condition anyway and in the case of the -3.7 and -4.2 deletions they are clinically silent when inherited alone. The whole area of how to approach antenatal counseling in populations with a high prevalence of multiple conditions is highly complex and the discussion of this element could be tightened up a bit.
A11) It is very difficult to carry on a proper ante-natal screening and counseling in our setting and we are fully aware of the limitations of our approach. At the beginning all women diagnosed with abnormal Hb were counseled. As explained already in A7 the first aim for genotyping subjects with low %HbA₂ was to rule out alpha thalassaemia carriage; when genotyping became available and the results showed that half of women did not have mutations on the alpha genes and another 40% were silent carriers, we modified the counseling accordingly. We are aware the discussion is quite long but we also wanted to touch on the several aspects that make this setting peculiar.

Minor
1) Though it’s an editorial decision, I would have assumed that since this is a UK based journal the spelling should be in UK English throughout.
We agree and have changed the spelling to UK English; more than an editorial choice, the first author is an Italian who has a relaxed approach towards spelling.
2) Abstract Methods: Blood groups were “diagnosed” makes it sound as if these are a disease! I would change to “determined” or similar.
We have cut this part.
3) “resource rich countries” – preferred current term is “high-income countries”
This has been changed
4) Final paragraph of the introduction could probably be condensed and moved to join the methods section. I would replace by a short sentence summarizing the aims of the current study
This has been done.
5) Please give the make and model of the HPLC machine used for the haemoglobin phenotyping.
We have added it.

Could do with a good proof-read throughout. A few examples I picked up:
1) Introduction line 3 “…living in area of past…” should read “…living in areas of past…”
2) Abstract Methods: Data were available rather than data was available (data are plural in this context).
3) page 4 line one column 2 – “SAE” should read “SEA”.
4) Table 6 – use the Greek symbol for alpha rather than the letter A.
5) page paragraph 2 left column “found to be carrier..” should read “found to be carriers..”

We have revised the whole manuscript and hopefully corrected all typos and incorrect terms.

**Competing Interests:** I have no competing interests